

## SHORT NOTE

# *IN VITRO* MATURATION OF BOVINE GRANULOSA CELLS FOR STEROID HORMONE PRODUCTION

## MADURACIÓN *IN VITRO* DE CÉLULAS DE LA GRANULOSA BOVINA PARA LA PRODUCCIÓN DE HORMONAS ESTEROIDES

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### ADDITIONAL KEYWORDS

Cell culture. Culture medium.

### PALABRAS CLAVE ADICIONALES

Cultivo celular. Medio de cultivo.

### SUMMARY

This study aimed to evaluate the effects of preincubated granulosa cells on *in vitro* maturation and steroid hormones production in cows. Granulosa cells from the ovaries were cultured using the modified M-199 with Earle's salts. 1% bovine serum albumin was added and 5% fetal calf serum. Also 0.8% of antibiotics was also included. Cultures were incubated at 37°C in a 95% air and 100% relative humidity. There were significant differences among values of progesterone and estradiol concentration obtained for levels of inclusion of FSH and LH. It can be concluded that there is necessity for supplementation of hormones in culture medium for bovine IVM.

### RESUMEN

Se evaluó el efecto de células de granulosa preincubadas sobre la maduración *in vitro* y producción de hormonas esteroideas en vacas. Las células de granulosa de los ovarios fueron cultivadas empleando el M-199 modificado con sales de Earle. Se añadió 1% de albúmina de suero bovino y 5% de suero fetal de ternero. También se incluyó un 0,8% de antibióticos. Los cultivos fueron incubados a 37°C con 95% de aire y 100% de humedad relativa. Se registraron diferencias significativas entre las concentraciones de progesterona y estradiol obtenidas con los niveles de inclusión de FSH y LH. Puede concluirse que es necesario suplementar con hormonas el medio de cultivo para IVM en bovinos.

### INTRODUCTION

The ovary of the cattle is a complex endocrine gland responsible for production of sex steroids and is the source of fertilizable ova for reproduction. The ovary possesses two primary steroidogenic cell types. The theca cells are responsible for androgen synthesis while the granulosa cells are responsible for conversion of androgens to estrogens as well as progesterone synthesis. These cells undergo a transformation in the luteal phase of the menstrual cycle, converting them from estrogen producing, to predominantly progesterone producing cells. Estradiol is produced by granulosa cell aromatisation of androgens secreted by the theca cells (Fortune, 1994). Estradiol synergises with the gonadotrophins to regulate the expression of FSH and LH receptors on granulosa cells, which are important developmental checkpoints in the lifespan of the follicle at recruitment and selection respectively (Dierich *et al.*, 1998, Ma *et al.*, 2004). Towards the end of the ovarian cycle, secretion of oestradiol by the dominant follicle stimulates the pituitary LH surge, which induces ovulation (Moenter *et al.*, 1990). Consequently, this study aimed to evaluate the effects of preincubated granulosa cells on steroid hormones production in bovine animals.

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## MATERIALS AND METHODS

This study was conducted at the Department of Animal Physiology, University of Agriculture, Abeokuta, Nigeria. Ten pairs of ovaries from cows were collected from local abattoirs and kept in normal saline for transport to the laboratory.

### PREPARATION OF CULTURE MEDIA

For granulosa cells culture, modified M-199 with Earle's salts was used. It was diluted in 1 litre of de-ionized water and 3.36 mM of  $\text{NaHCO}_3$  was added to the 1 litre TCM-199 solution; 1% BSA was then added to half of the TCM-199 while 5% FCS was added to 47.5 ml of TCM-199; 0.8% of antibiotics was again added to the medium after which 0.2  $\mu\text{m}$  Supor® membrane filter was used to filter the media.

### METHODOLOGY

In the laboratory, the ten pairs of ovaries were washed in fresh sterile saline. Ovaries were dried before the granulosa cells recovery from the follicles. All visible follicles were aspirated using 18" needle. Granulosa cells were separated by centrifuging at 3000 rpm for 10 min. Cell pellet were then mixed thoroughly with the culture

medium and aliquoted at 50  $\mu\text{l/ml}$  per well into a 24-well micro plates.

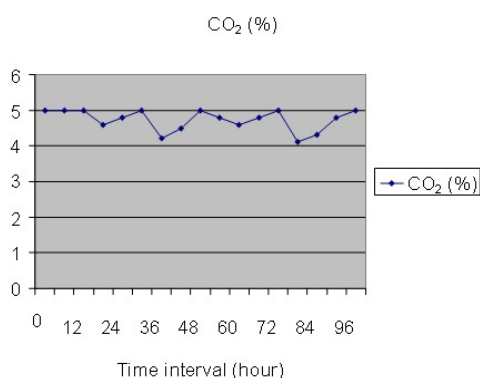
Cultures were incubated at 37°C in a 95% air and 100% RH. The graphical representation data of  $\text{CO}_2$  and RH are presented in **figures 1 and 2**. Medium was changed after 2 days of culture and to obtain optimal attachment, cells were maintained in the presence of 10% FCS for the first 48 hours of culture. Hormonal treatments were maintained for 2 days while on the 3<sup>rd</sup> day, testosterone (10 ng/ml) was added as substrate for estradiol production. The experiment was terminated on the 4<sup>th</sup> day.

### DATA COLLECTION

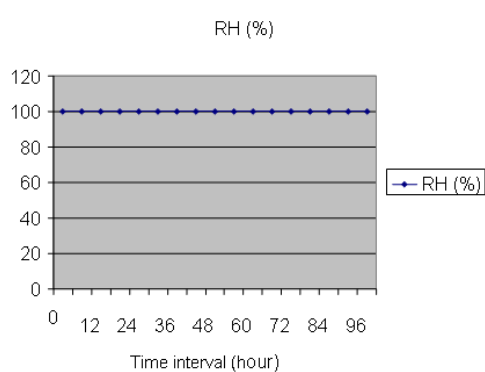
At the termination of the experiment, culture medium samples were collected for determination of progesterone and estradiol concentrations by Radioimmunoassay method. The design of the experiment includes: (1) additives supplementation i.e. FSH and LH and (2) inclusion levels: (a) 0 ng/ml (b) 0.1 ng/ml (c) 1 ng/ml (d) 10 ng/ml and (f) 100 ng/ml with each treatment replicated 3 times.

### STATISTICAL ANALYSIS

Data collected were subjected to analysis of variance. Significant means were



**Figure 1.** Graphical representation of  $\text{CO}_2$  regime in the inner chamber of  $\text{CO}_2$  incubator. (Representación gráfica del régimen de  $\text{CO}_2$ , en la cámara interior del incubador de  $\text{CO}_2$ ).



**Figure 2.** Graphical representation of relative humidity regime in the inner chamber of  $\text{CO}_2$  incubator. (Representación gráfica del régimen de HR, en la cámara interior del incubador de  $\text{CO}_2$ ).

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separated using Duncan Multiple Range Test. All data were analyzed using Systat Analytical Computer Package.

### RESULTS

In relation with the effects of FSH on bovine granulosa cells culture *in vitro* for the production of progesterone and estradiol, there was significant difference among the values of progesterone concentration obtained for different levels of inclusion of FSH (**table I**). The addition of 10 ng/ml of FSH had the highest value of progesterone. Also there were no significant differences between this value and the progesterone values obtained with other inclusion levels. Estradiol values followed the same pattern of result obtained in the progesterone.

The effects of LH on bovine granulosa cells culture *in vitro* for production of progesterone and estradiol are presented in **table II**. From the table, progesterone values recorded the highest at 100 ng/ml. Although this value was not significantly different from those recorded at 50 ng/ml and 10 ng/ml LH inclusion levels.

**Table I.** Effects of FSH on bovine granulosa cells culture for progesterone and estradiol production *in vitro*. (Efectos de la FSH sobre el cultivo de células de granulosa bovina sobre la producción *in vitro* de progesterona y estradiol).

FSH inclusion level (ng/ml)	Progesterone (ng/ml)	Estradiol 17 $\beta$ (pg/ml)
0	0.88 <sup>d</sup>	1.41 <sup>d</sup>
0.1	1.41 <sup>c</sup>	2.05 <sup>c</sup>
1.0	2.24 <sup>b</sup>	3.11 <sup>b</sup>
10	2.95 <sup>a</sup>	5.81 <sup>a</sup>
50	2.94 <sup>a</sup>	5.69 <sup>a</sup>
100	2.82 <sup>a</sup>	5.62 <sup>a</sup>
S.E.M.	0.11	0.20

<sup>abcd</sup>means in the column with different superscript are significant ( $p < 0.05$ ).

### DISCUSSION

Primary cultures of bovine granulosa cells have previously been used to examine a number of events (Amer *et al.*, 2008; Yang and Rajamahendhran, 1998). From the results presented above, there were significant effects of FSH on the progesterone and estradiol production measured and this is in line with the results of Mao *et al.*, 2002). It was observed in this research that an increase in the level of FSH from 10 ng/ml upward generates no significant increase in the progesterone production. The inclusion of LH also had significant effects on progesterone and estradiol production. This is a pointer that granulosa cell proliferation is a measure of steroidogenic capacity of cells. Meanwhile, both progesterone and estradiol production reached the highest values when LH was included at 100 ng/ml in the maturation medium, therefore further increase in the level of inclusion of LH resulted in no corresponding increase in progesterone and estradiol production. The results obtained in this study is in line with Zuelke and Brackett, 1990 and Saeki *et al.*, 1991. In

**Table II.** Effects of LH on bovine granulosa cells for progesterone and estradiol production *in vitro*. (Efectos de la LH sobre células de granulosa bovina para la producción *in vitro* de progesterona y estradiol).

LH inclusion level (ng/ml)	Progesterone (ng/ml)	Estradiol 17 $\beta$ (pg/ml)
0	0.71 <sup>c</sup>	1.38 <sup>c</sup>
0.1	0.96 <sup>c</sup>	1.72 <sup>c</sup>
1.0	1.29 <sup>b</sup>	2.69 <sup>b</sup>
10	1.77 <sup>a</sup>	3.54 <sup>a</sup>
50	1.88 <sup>a</sup>	3.68 <sup>a</sup>
100	1.99 <sup>a</sup>	3.27 <sup>a</sup>
S.E.M.	0.08	1.14

<sup>abc</sup>means in the column with different superscript are significant ( $p < 0.05$ ).

conclusion, there is necessity for supplementation of hormones into culture medium for successful bovine *in vitro* maturation and it is suggested further that studies using abattoir ovaries may not be truly representative of the potential of the bovine ovary because mostly aged and underfed animals are slaughtered. Therefore, further studies should be focused on oocytes recovered from animals in good body condition.

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